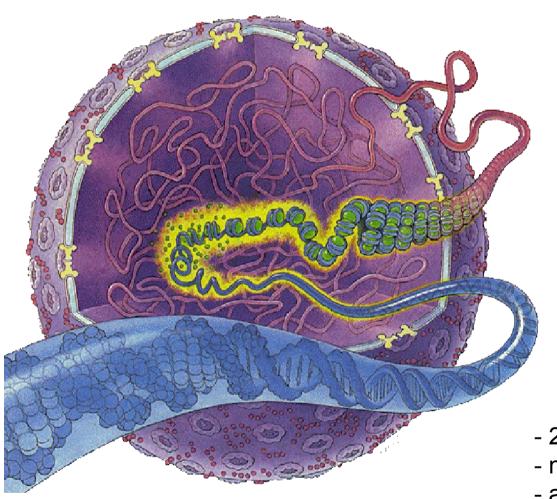
The human genome

A human cell nucleus



- 2 m DNA
- nucleus ≈ 10 μm diameter
- about 30 000 genes
- -10 000 different nuclear proteins

2 nm short region of DNA double helix 11 nm "beads-on-a-string" form of chromatin 30-nm chromatin 30 nm fiber of packed nucleosomes section of chromosome in an extended form condensed section of chromosome 700 nm centromere entire 1400 nm ayay haddid mitotic chromosome

from DNA to chromosomes

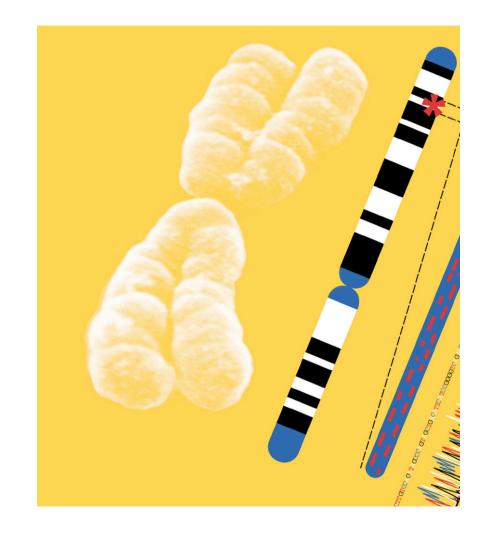
Telomeres

centromere

p arm

q arm

sister chromatids



How many genes are there in the human genome?

30,000 more? or less?

What percentage of the genome actually codes for genes?

only about 1-3% What is the rest for?

How large is the human genome?

3 billion base pairs

The largest gene: dystrophin (associated with Duchene's muscular dystrophy)

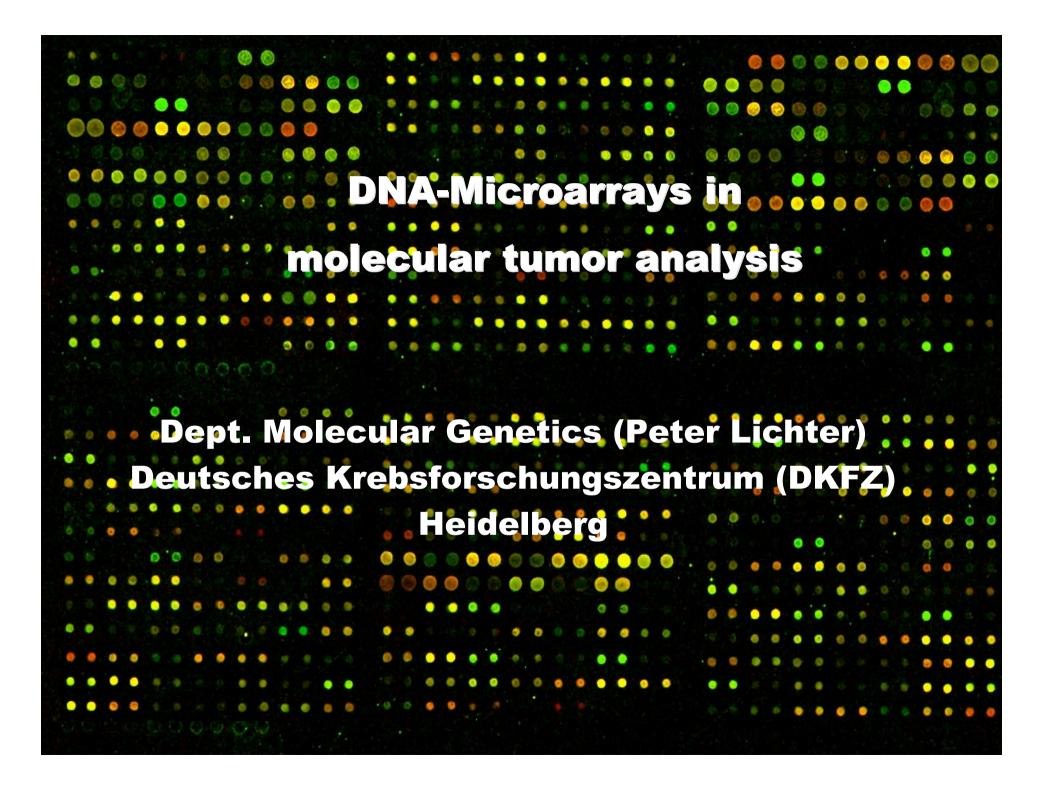
2 400 000 bases

The smallest genes: tRNA genes, about 100 bases

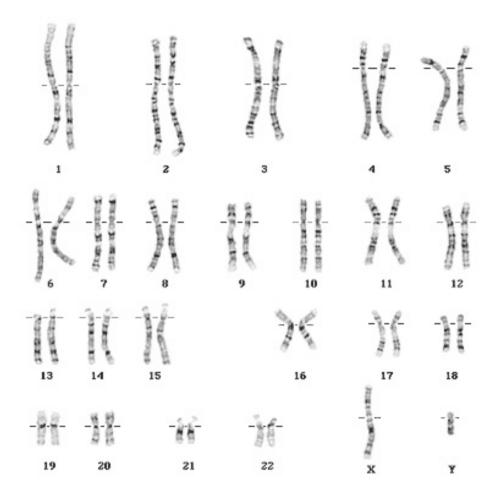
How did they estimate the number of genes?

- 1) genomic sequencing extrapolation from sequencing large chromosome regions.
- 2) CpG island numbers (short stretches of DNA--about 1-2 kilobases long) About 56% of genes are associated with CpG islands. The total number of CpG islands is about 45000.
- 3) EST=expressed sequence tags

Nucleotide content of Human DNA

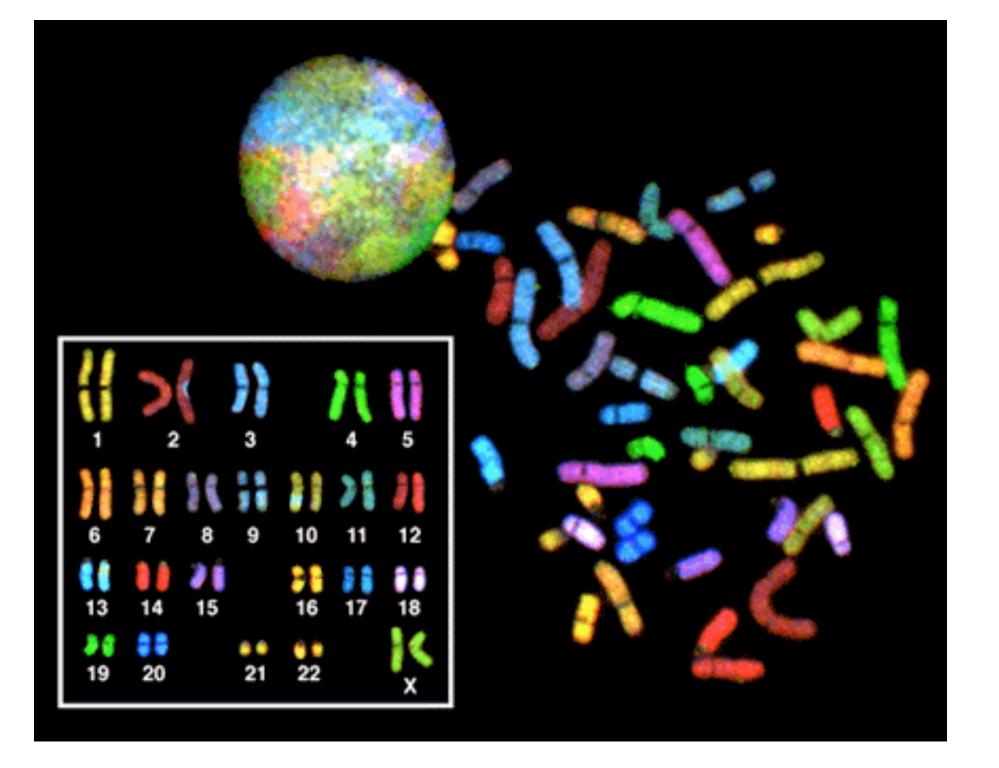


Human chromosomes



24 chromosomes total

- 22 autosomal chromosomes
- X, Y

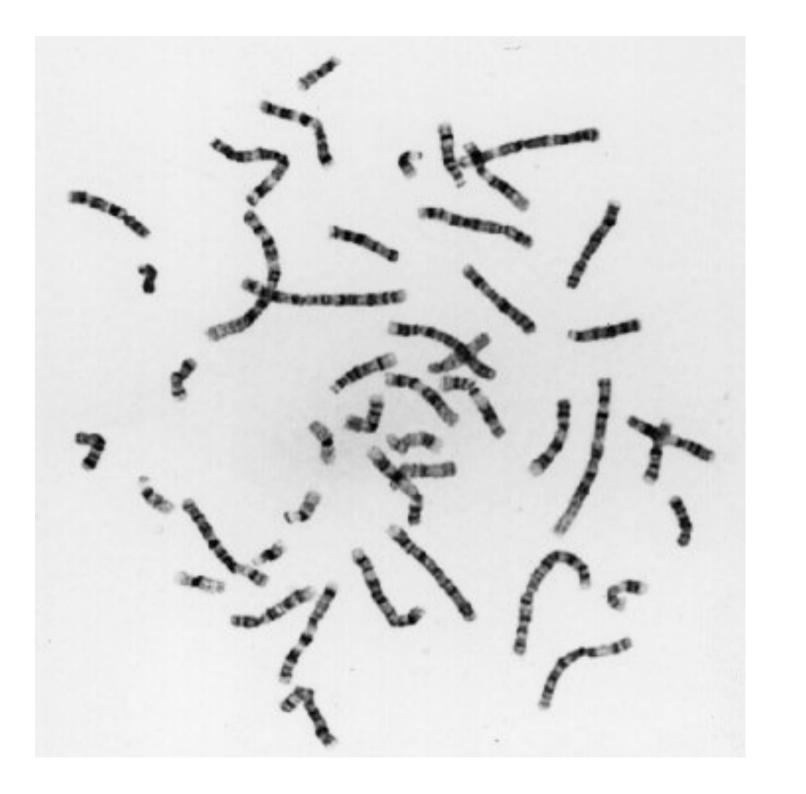


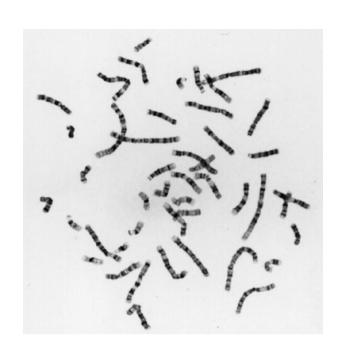
The 22 somatic chromosomes and the X and Y chromosomes are easily identified by

- -size
- -centromere location
- -secondary constrictions (present on the long arms of 1,9, and 16)
- -G band patterns

19 and 22 are gene rich

4 and 18 are gene poor





Properties of the Dark G Bands

AT rich
Dnase insensitive
condense early in the cell
cycle but, replicate late
gene poor
but, genes that are there,
have large introns

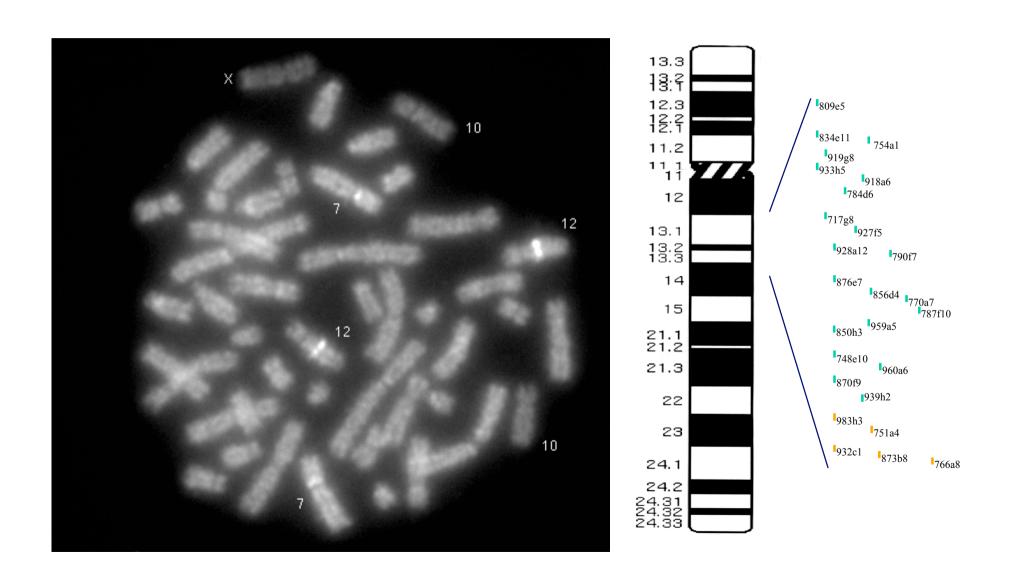
More than 99.9% of human DNA sequences are the same across the population of all humans in the world.

Single-nucleotide polymorphisms (SNPs) occur about once every 100 to 300 bases.

polymorphism = many forms

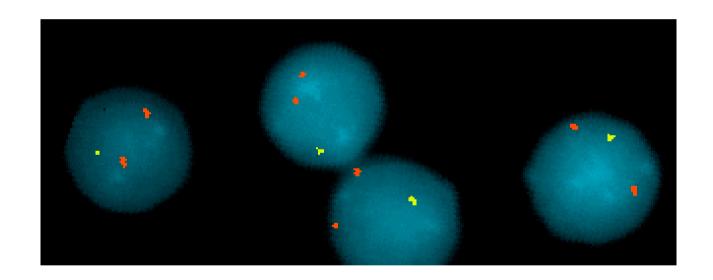
Amplification of oncogenes in a human tumour

gene card



B-cell chronic lymphocytic leukemia (B-CLL)

mature but immuno-incompetent B-cells in peripheral blood Impaired proliferation ?
Impaired induction of apoptosis



Profile of genomic alterations:

Loss: subregions within 6q, 10q, 11q (ATM), 13q (BCMS?), 17p (TP53)

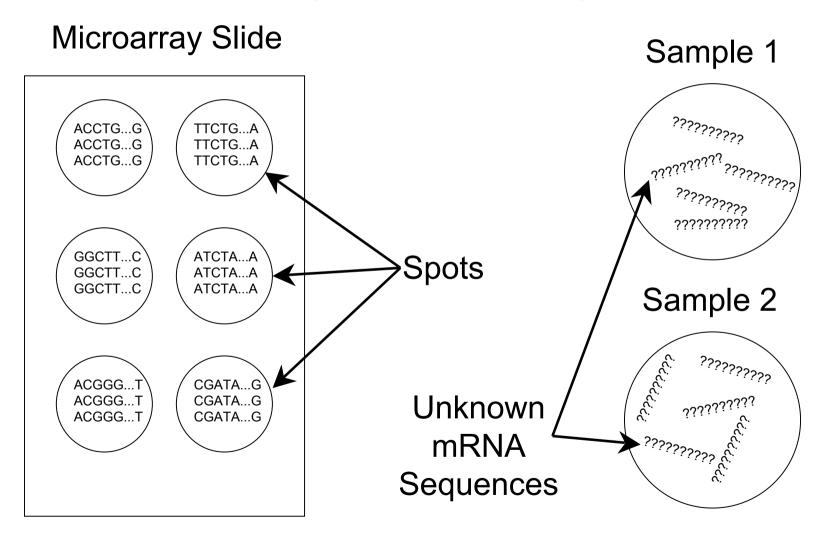
Gain: subregions within 3q, 8q (MYC), 12q

DNA microarrays

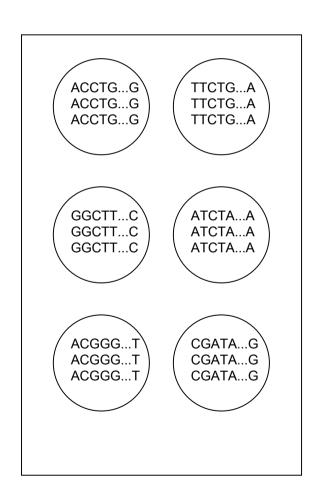
Microarray Technology

- Microarrays allow researchers to measure the abundance of thousands of mRNA transcripts in multiple biological samples.
- By understanding how transcript abundance changes across experimental conditions, researchers gain clues about gene function and learn how genes work together to carry out biological processes.

Two-Color Microarrays (cartoon version)



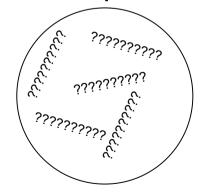
Extract mRNA



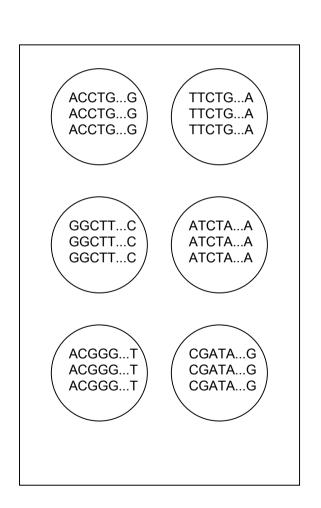
Sample 1

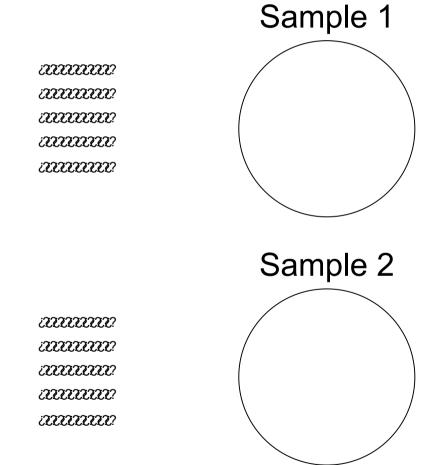


Sample 2

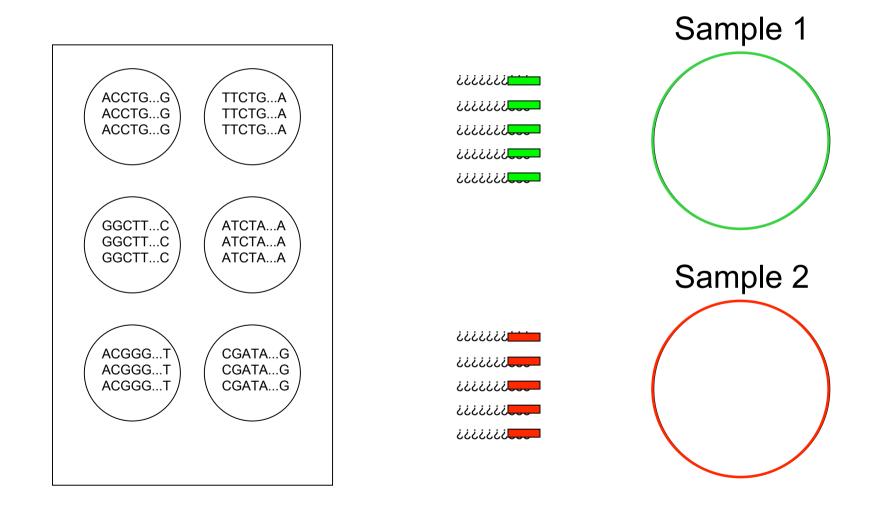


Convert to cDNA





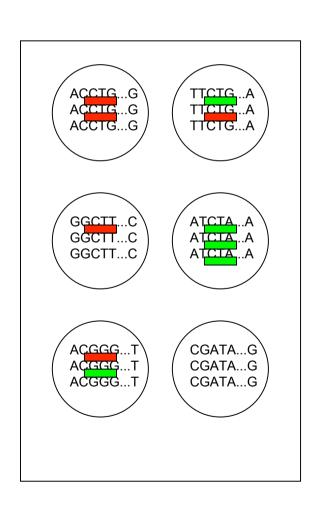
Tag cDNA from Different Samples with Different Fluorescent Dye

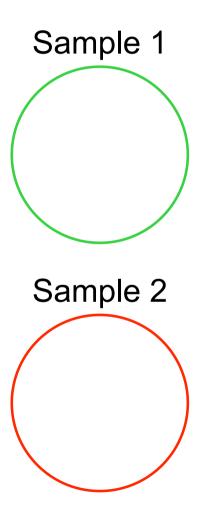


Hybridize cDNA to the Slide

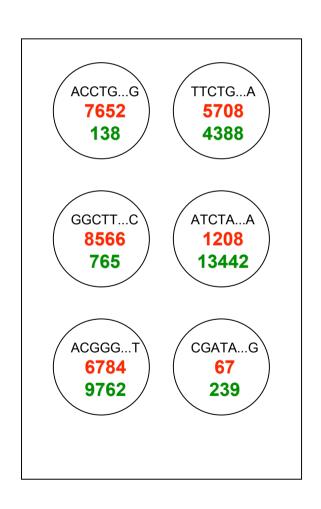


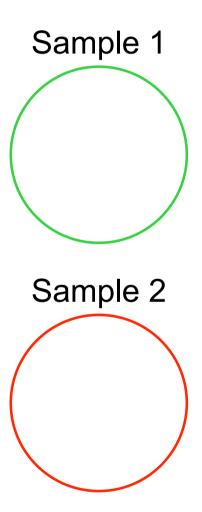
Hybridize cDNA to the Slide





Scan and Quantify Signals

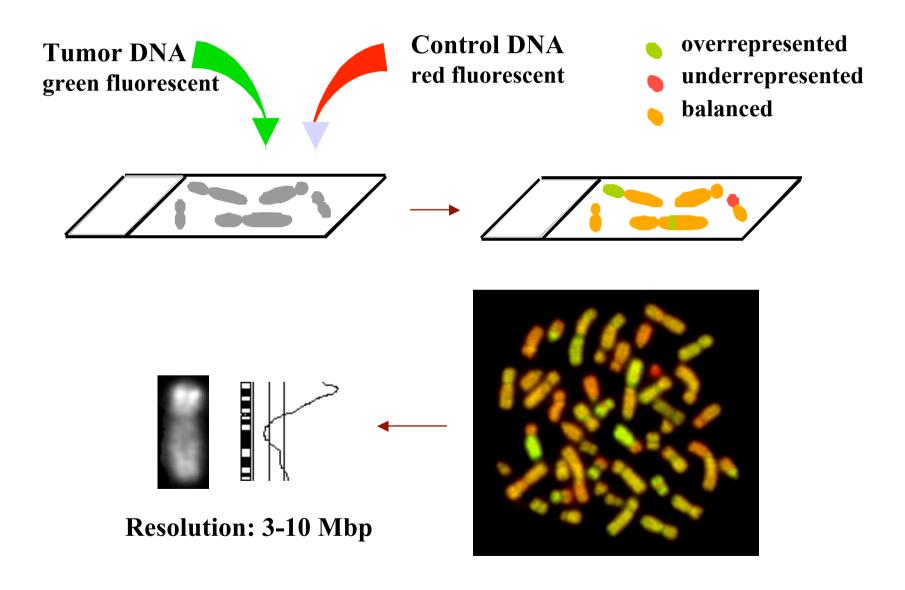




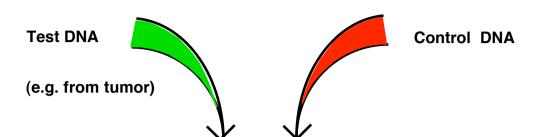
Matrix-CGH

Matrix-CGH is a chip-based, high-resolution method for the analysis of genomic-DNA copy number. It is used this important tool to characterize chromosomal aberrations in human tumors.

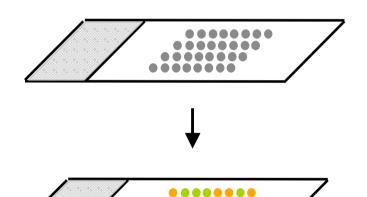
Comparative Genomic Hybridization (CGH)



Matrix-CGH



Solinas-Toldo et al. Genes Chromosom. & Cancer 20, 399-407, 1997

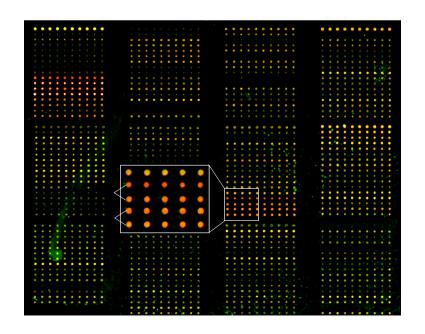


Matrix of microarrayed genomic DNA fragments

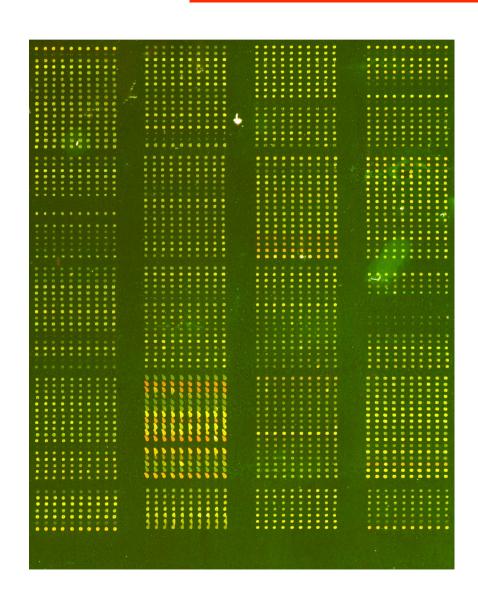


= material lost in test genome

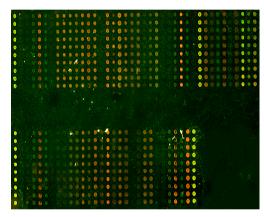
= material balanced in test genome



Diagnostic DNA-chips:

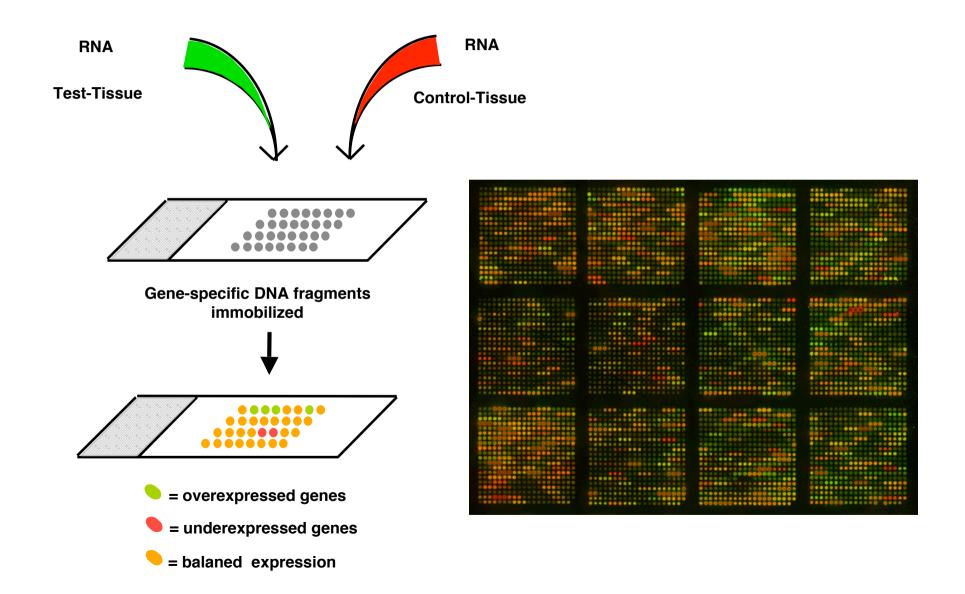


e.g. diagnosis of B-cell chronic lymphocytic leukemia

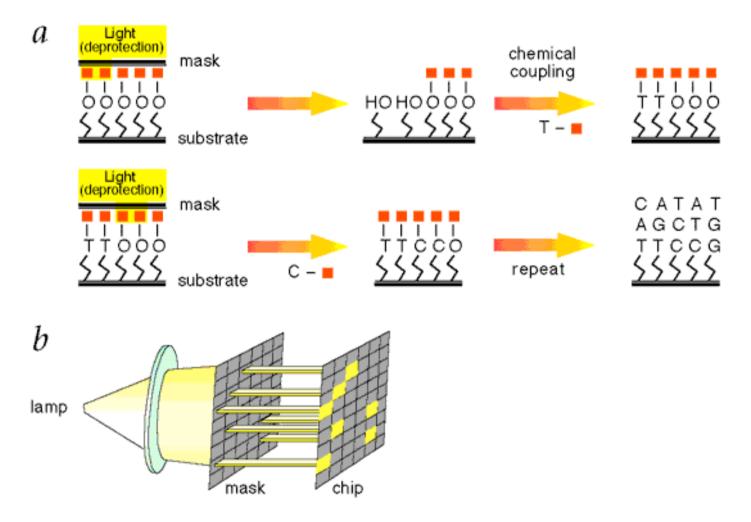




Gene Expression Profiling

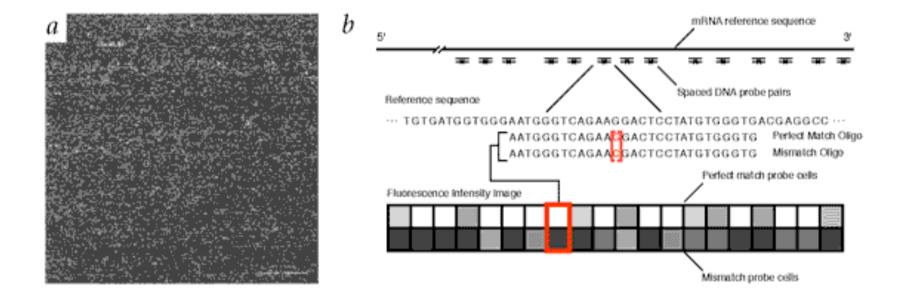


Affymetrix-Chips and CYP450



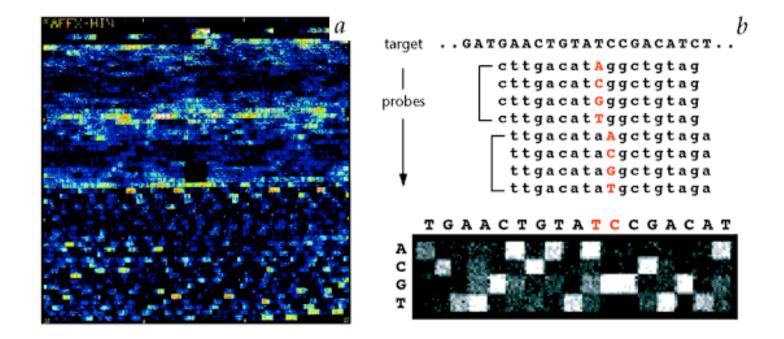
Light directed oligonucleotide synthesis.

A solid support is derivatized with a covalent linker molecule terminated with a photolabile protecting group. Light is directed through a mask to deprotect and activate selected sites, and protected nucleotides couple to the activated sites. The process is repeated, activating different sets of sites and coupling different bases allowing arbitrary DNA probes to be constructed at each site. **b**, Schematic representation of the lamp, mask and array.



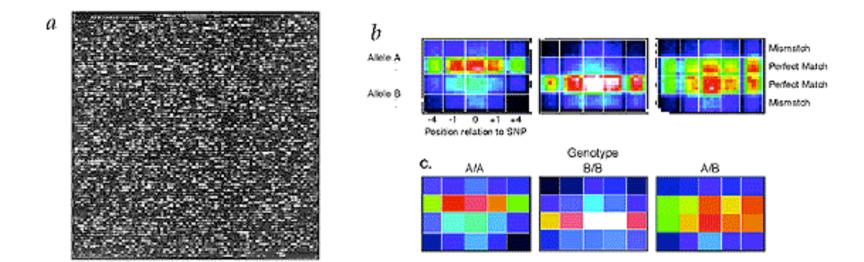
Gene expression monitoring with oligonucleotide arrays.

a, A single 1.28 1.28 cm array containing probe sets for approximately 40,000 human genes and ESTs. This array contains features smaller than 22 22 m and only four probe pairs per gene or EST. **b**, Expression probe and array design. Oligonucleotide probes are chosen based on uniqueness criteria and composition design rules. For eukaryotic organisms, probes are chosen typically from the 3' end of the gene or transcript (nearer to the poly(A) tail) to reduce problems that may arise from the use of partially degraded mRNA. The use of the PM minus MM differences averaged across a set of probes greatly reduces the contribution of background and cross—hybridization and increases the quantitative accuracy and reproducibility of the measurements.



Sequence analysis arrays.

a, Image of an HIV-1 genotyping array (HIV PRT) hybridized to labelled PCR-generated DNA copies of the protease (PR) and reverse transcriptase (RT) genes (0.8 0.8 cm array). Each base on both strands covering a total of 1515 base pairs is interrogated in the HIV-1 PR (codons 1–99) and RT (codons 1–400) genes. Additionally, to ensure detection in the event of multiple mutations occurring in close proximity, most common drug-resistance conferring mutations are encoded on the array as specialized tilings. b, General tiling strategy. Detection of mutations or polymorphisms in a sequence is accomplished by using a four-probe interrogation strategy. In this illustration, four 17-mer oligonucleotide probes are used to determine the identity of the base in the middle of the probe sequence. The probe that forms the most stable duplex will provide the highest fluorescent signal among the four probes assigned to interrogate the central base. The next nucleotide in the target sequence is interrogated in the same manner, using another set of four oligonucleotide probes. Probes with interrogation positions other than the central position, or probes of different lengths can also be used to query the targeted base. Analysis of both strands of a target can be carried out on the same array to increase the confidence of the base determination.



Genotyping arrays.

a, A single array with over 120,000 probes designed to determine the genotype of a sample at over 3,000 biallelic loci. **b**, The fluorescence intensity pattern for a set of probes designed to interrogate a single locus showing the presence of an AA homozygote, a BB heterozygote, and a BB homozygote. The upper and lower halves of the probe blocks interrogate the A and B alleles, respectively. Each half consists of pairs of probes centered on the polymorphic position and offset one and four bases to either side. The probe pairs consist of a perfect match and single base mismatch to the reference sequence for the specific allele. For each locus, interrogation blocks are included for both the sense and anti–sense strands.

Table 2 • Commercial GeneChip® probe arraysa

Application	Species	Information	
expression	human ^b mouse ^b rat ^c yeast (S. cerevisiae) Drosophila melanogaster ^c Arabidopsis thaliana ^c C. elegans ^c E. coli ^c other bacteria ^d targeted ^e custom ^f	~42,000 genes/ESTs ~30,000 genes/ESTs >11,000 genes/ESTs whole genome (all ORFs) >12,000 genes/ESTs >12,000 genes/ESTs whole genome (all ORFs) whole genome (all ORFs) whole genome (all ORFs) functionally selected gene sets any eukaryotic organism	
genotyping	human	~2,000 SNPs	
polymorphism screening	human	screening service	
variant analysis	human human HIV-1, Clade B	CYP450 (2D6, 2C19) p53 (exons 2–11) HIV (protease, rev. transcriptase)	

The GeneChip® System is required to run the arrays. The complete system is priced at \$175,000 in the U.S. and includes the GeneChip® Fluidics Station, Hewlett Packard GeneArray™ Scanner, GeneChip® workstation and GeneChip® 3.1 analysis software.

bArray update including additional gene and EST information expected in 1999. Expected to be available mid-1999. Five bacterial species planned for late 1999. Specific subsets of genes selected based on their relevance to a given biological, clinical or disease area. Gene or EST sets (public or proprietary) chosen by external users.

Table 3 • Array capacity and feature size

Feature Size	Expressiona	Sequence Analysisb	Genotyping ^c
50 μm	1600_6400 genes	8–16 kb	2,000–4,000 markers
20 μm	10,000_50,000 genes	50–100 kb	12,000–25,000 markers
2 μm	>1 million genes	500–1,000 kb	1.2–2.5 x106 markers

All numbers calculated for 1.28x1.28 cm arrays. ^aAssuming 4–20 probe pairs per gene. ^bAssuming 4–8 probes per basepair. ^cAssuming 6–32 probes per marker.

AmpliChip CYP450

komplexe und umfassende Analyse von

29 CYP2D6-Varianten sowie
2 CYP2C19-Varianten
in einem einzigen Assay



Roche Diagnostics: erste Firma, welche die Affymetrix Mikroarray-Technologie für eine diagnostische Fragestellung nutzt

CYP2D6-kodierte Enzyme metabolisieren:

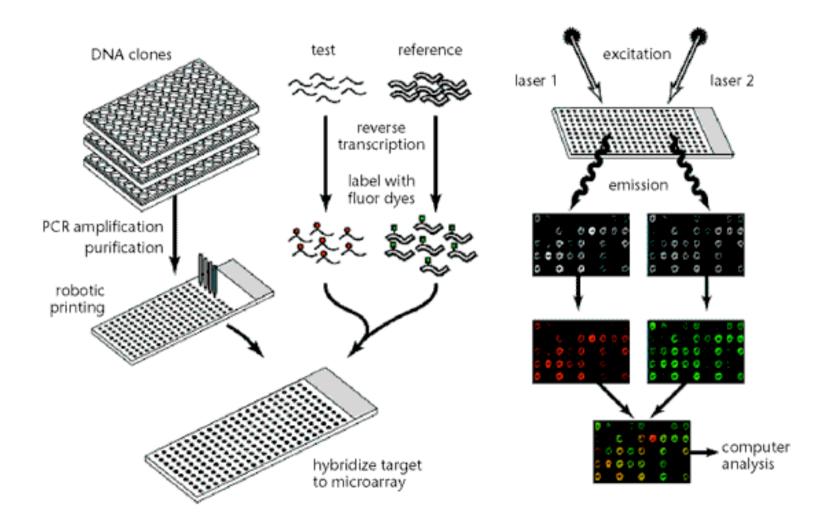
- zahlreiche Anti-Depresiva
 - Anti-Psychotika
 - Anti-Arrhytmika
 - Schmerzmittel
 - beta-Blocker

CYP2C19-kodierte Enzyme:

metabolisieren diverse Substanzen, u.a. Protonenpumpen-Inhibitoren, Anti-Koagulantien, Benzodiazepine, Anti-Malariamittel

Making microarrays

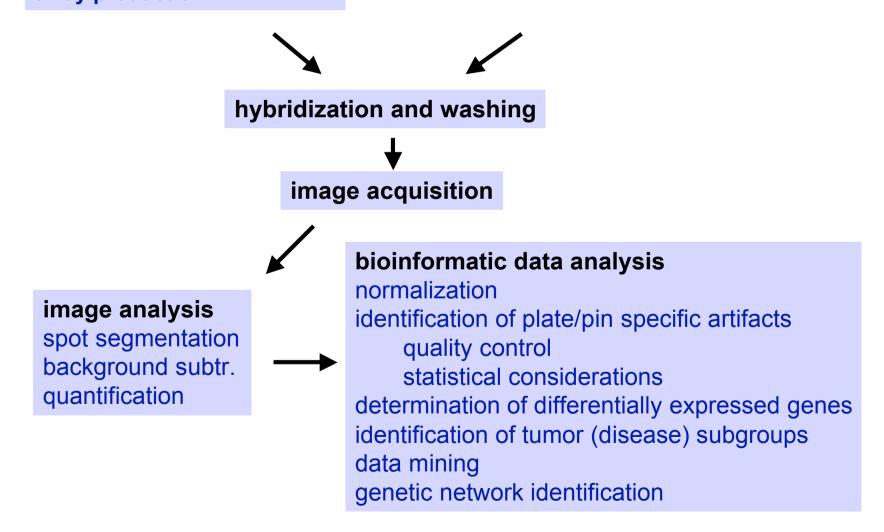
The Principle of cDNA Microarrays



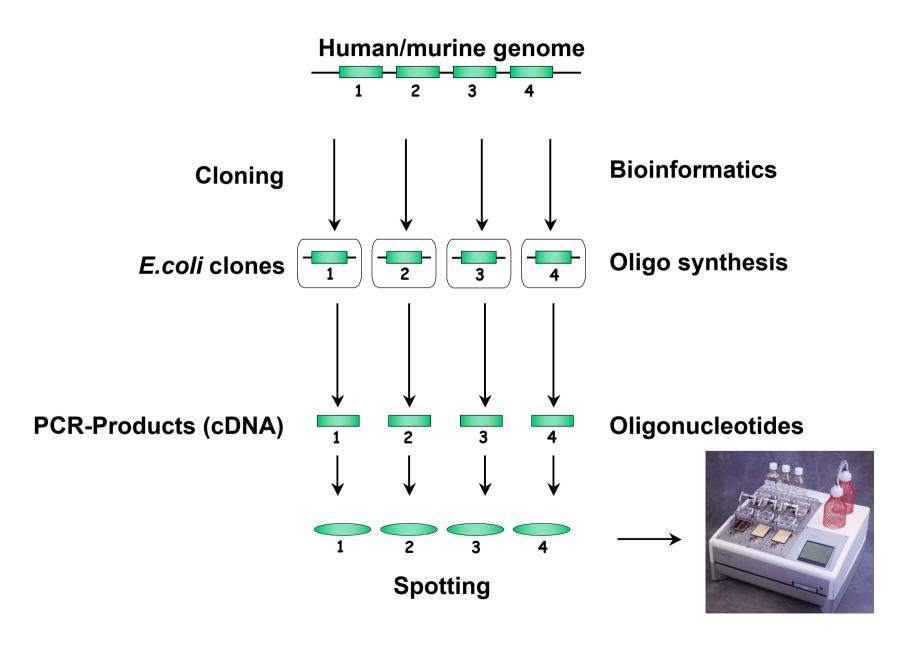
Duggan DJ, Bittner M, Chen Y, Meltzer P, Trent JM. Expression profiling using cDNA microarrays. *Nat Genet*. 1999, 21(1 Suppl):10-14.

cDNA array construction clone validation PCR amplification array production

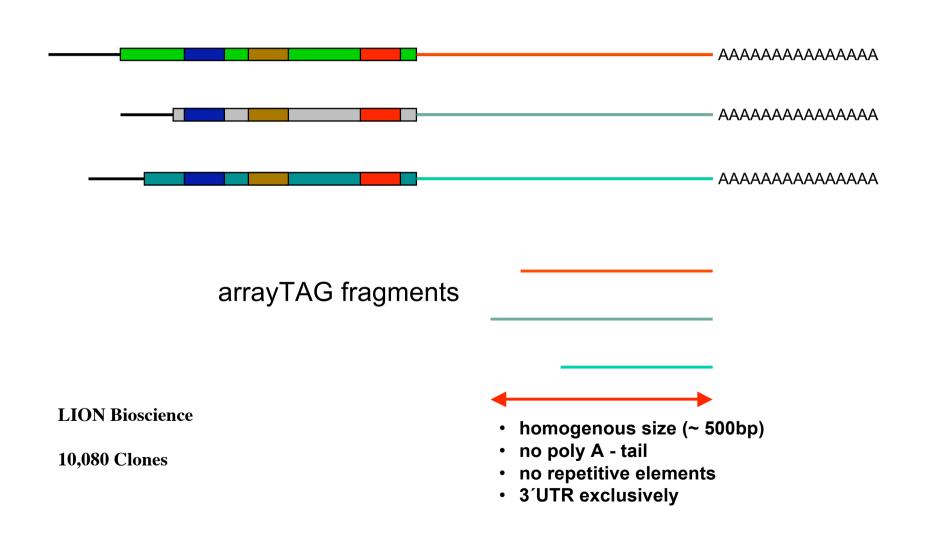
sample isolation & labeling reverse transcription of RNA using CY3/CY5 for labeling



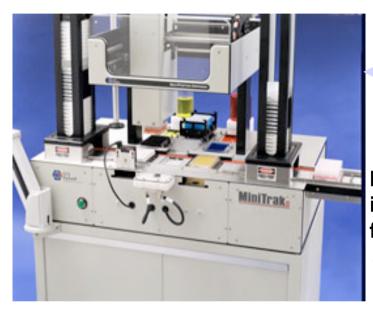
Expression Profiling



arrayTAGs: Optimized cDNA Fragments for Microarrays



The Liquid Handling System I

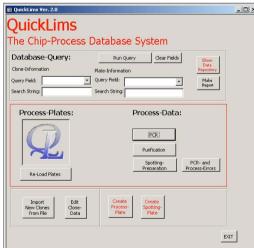


Read Barcode





Request plate information from QL



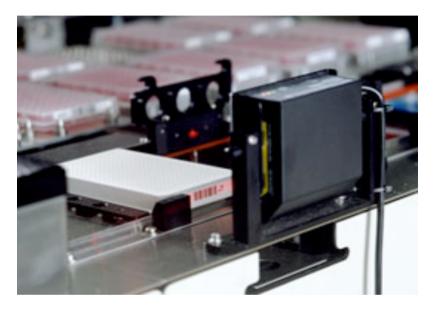


Pipet according to step information (provided by QL)

The Liquid Handling System II



Packard Bioscience MiniTrak conveyor-based robotic liquid handling system



The robot uses a bar code reader to identify individual process plates. This enables the VB control script, which is connected to a custom database system, to monitor, automate and document the whole production process

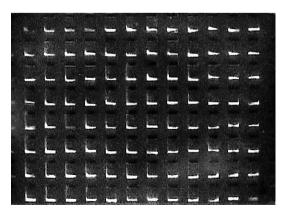
PCR Process



MiniTrak Liquid Handling System



96 Well Steel Replicator



96-well Ready-to-Run Agarose Gel

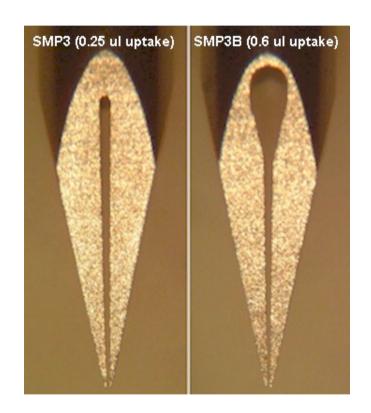


4 x 4 Block PCR Machine (96 Wells)

Production of cDNA Microarrays I



GeneMachines OmniGrid multiaxis microarrayer

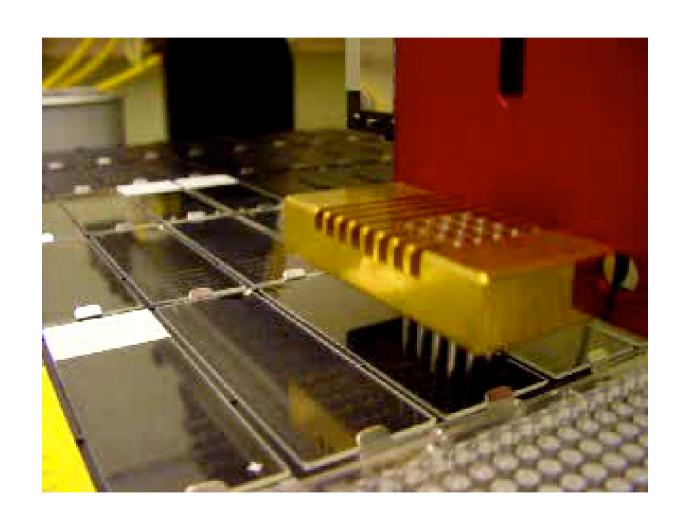


Telechem Stealth Micro Spotting Pins SMP3 and SMP3B (split / capillary pins)

Production of cDNA Microarrays II

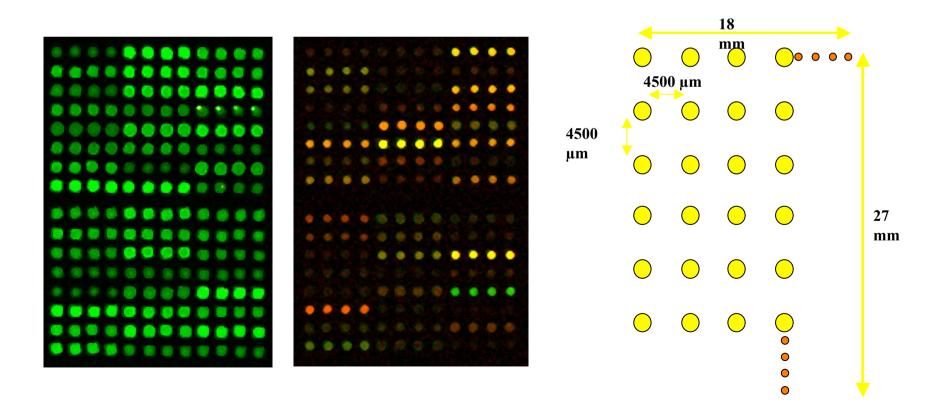


Production of cDNA Microarrays III



Spots at 140 µm Dot Spacing

- Spotting can be performed at 140 µm dot spacing
- Printable area (for HybStation): 18 x 54 mm
- $(4500 : 140)^2 \times 4 \times 6 \times 1 \times 2 = 49,590 \text{ spots}$
- 24,900 clones as replicate array, 24 pins (Virtek Microarrayer)



Hybridization of Microarrays



TeleChem hybridization cassette



Genomic Solutions GeneTAC Hyb-Station (ASP)

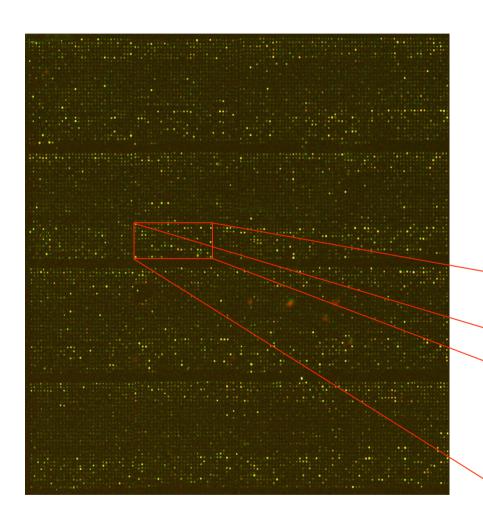
Scanning of Microarrays



Axon GenePix 4000A Microarray Scanner

- Resolution: 10 µm / pixel
- 2 laser-PMT units
- Detection @ 532 nm (Cy3) and 635 nm (Cy5)
- Speed: 3-4 min /slide

What it looks like...



NIA 15K hybridized with 20 µg RNA of 3T3 and F9 cells, respectively

